Laboratory Animals (Injections, Marking, and Mice Anatomy)

- A laboratory model animal refers to any experimental animal species that is used in research and drug development processes to simulate human anatomy, physiology, and diseases.
- These model animals play a crucial role in understanding basic biological mechanisms, assessing drug efficacy and safety, and developing new treatment methods.
- The aim of injection: To form protective antibodies in the serum and these antibodies are preserved by memory cells.



Classification of experimental animals

- ✓ **Rodents** (Mice, Rats, Guinea pigs, Hamster... etc.)
- ✓ Non Rodents (Rabbit, Dog, Cat, Monkey.....etc.)



Characteristic of lab Animals use in injection:

- 1. Empty of infection (healthy)
- 2. Specific weight, age and type
- **3.** Genetically pure

- To study immunological and pathological process of disease.
- For production of antibodies.
- To study the antigenicity of vaccines.
- Source of Blood (erythrocytes , serum , plasma).

Injections and blood collecting methods in the laboratory animals

- Intra muscular injection.
- Intra cerebral injection
- Intra venous injection.
- Intra peritoneal injection.
- Intra dermal Injection.
- Intra nasal injection.
- Subcutaneous injection.
- Intra foot-pad injection.
- Intra thymic injection.

Conditions of injections in laboratory animals:

- 1. Sterile cleaned syringe.
- **2.** Clean area by alcohol 70%.
- **3.** No bubbles in the syringe.

Injection in rodent animals:

- Intra muscular injection.
- Intra dermal Injection.
- Intra peritoneal injection.
- Subcutaneous injection

Injection in non- rodent animals:

- Intra muscular injection.
- Intra venous injection.
- Intra peritoneal injection.
- Intra dermal Injection.
- Subcutaneous injection



Blood collecting in the laboratory animals:

Collection blood from laboratory animals is frequently necessary for a variety of experimental uses such as antibody production.

Type of Blood collecting:

- Cardiac puncture
- Orbital puncture
- Orbital plexus
- Orbital venous sinus
- The tail
- Central auricular artery

Blood collecting in rodent animals:

- Orbital plexus.
- Cardiac puncture.
- The tail.

Blood collecting in non- rodent animals:

- Central auricular artery.
- Cardiac puncture

Marking of animals

Temporary marking:

- 1. Cage marking.
- 2. Dye marking.
- 3. Hair clipping.
- 4. Neck bands

Permanent marking:

- 1. Natural marking.
- 2. Ear punching.
- 3. Toes clipping.
- 4. Brand



Mice Anatomy (Field Mouse Dissection)

The field mouse is a small mammal that has many anatomical structures that are similar to human structures, however much smaller because the mouse is a mammal like humans, it is warm blooded and has a four---chambered heart.

Steps for Dissecting a Mouse:

1. Preparation

- Gather necessary tools: scissors, forceps, scalpel, dissection board, gloves.
- Ensure a clean and safe working environment.

2. Positioning the Mouse

- Place the mouse on the board, securing the legs with tape if needed.

3. Opening the Abdominal Cavity

- Use the scalpel to make a small incision in the abdominal area.
- Gently widen the incision to access internal organs.

4. Examining the Organs

- Liver: Identify its location and examine its size and color.
- Lungs: Observe them and ensure they are healthy.
- Heart: Check it by making an incision in the chest area.
- Stomach and Intestines: Examine their location and condition.

5. Inspecting the Nervous System

- Carefully remove the skull to examine the brain.
- Identify different areas of the brain.

6. Cleaning the Area

- Use water or saline solution to clean the internal organs if necessary.

7. Documentation

- Record observations about each organ and note any abnormalities.

8. Re-dissecting (if necessary)

- If needed, further examinations can be performed or tissue samples can be taken.

9. Disposing of Waste

- Dispose of the mouse and dissection materials safely according to health guidelines.



