

# PREPARATION, CHARACTERIZATION AND ANTIMICROBIAL ACTIVITY OF POLYVINYL ALCOHOL/ POLYVINYL PYRROLIDONE/ CHITOSAN NANO COMPOSITE

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**ABSTRACT :** Ternary polymer blend of chitosan/poly vinyl alcohol/ poly vinyl pyrrolidone was prepared by solution casting method, nanocomposite was prepared by sonication method with nano Ag and Zn. All prepared compounds have been characterized by FT-IR, SEM, DSC, as well as Biological activity. Antimicrobial activity related to prepared blends and Nanocomposites against six types of bacteria namely, *Staphylococcus aureas*, *E. faecalis*, *S.typhi*, *P. aeruginosa*, *Bacillus subtilis*, *Escherichia coli* and *C. albicans* fungal were examined and evaluated. The results reveal that the prepared polymer blends and nanocomposites have good antimicrobial activity against all kinds of microbials.

**Key words :** Chitosan, PVA, PVP, polymer blend, nanocomposite, antimicrobial polymer.

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## INTRODUCTION

Bionanocomposites are made up of a biopolymer matrix (continuous phase) and a reinforcing agent (dispersed phase) made up of particles with sizes ranging from 1 to 100 nm (Joseph *et al*, 2020). Most of the biopolymers are inherently weak in physical, mechanical and thermal attributes compared to widely-used petro-based polymer (Garavand *et al*, 2020). Nanobiocomposites possess a couple of advantages over the biocomposites and even composites; in turn, loading nanofillers into the biocomposites could improve their thermal, structural, mechanical and barrier attributes in a safe way with widespread range of uses (Wei *et al*, 2020; Rezkita *et al*, 2020).

Chitosan is a polycationic linear polysaccharide produced from chitin that is naturally polycationic. Chitosan's poor solubility in neutral and alkaline solutions restricts its use. Chemical modification of composites or hydrogels, on the other hand, confers new functional characteristics for a variety of applications. Because of its low allergenicity, biodegradability, biocompatibility and non-toxicity, chitosan is regarded as a versatile biomaterial (Kumar *et al*, 2020; Cheung *et al*, 2015). Chitosan's unique properties enable it to be utilized in a

wide range of applications. Other biological characteristics of chitosan have been identified, including anticancer, antibacterial, and antioxidant capabilities (Aranaz *et al*, 2009). Chitosan has many approved biological properties. It is available in nature and can be fabricated into many forms like film, fiber, bead and powder (Jou *et al*, 2011). Despite the fact that Chitosan is not predominantly employed as an antibacterial agent, its usage as a component in food and pharmaceutical formulations has recently increased, and the pharmacological actions of this versatile carbohydrate have begun to emerge. Understanding the many variables that influence its antimicrobial action, on the other hand, has become a critical problem for improved chitosan formulation use and optimization (Raafat *et al*, 2009; Martins *et al*, 2014; Kassem *et al*, 2019). Chitosan has been recognized as a potential antifungal substance (Qin *et al*, 2020).

PVA (polyvinyl alcohol) is a non-toxic, water-soluble synthetic polymer with strong physical and chemical characteristics as well as the capacity to create films (Parida *et al*, 2011). Because of their permeability, biocompatibility and biodegradability, chemically crosslinked PVA hydrogels are gaining popularity in biomedical and biochemical applications (Aytimur *et al*,

2013). PVA is a kind of versatile polymer and contains high mechanical applications (Arefian *et al*, 2020).

PVP (polyvinyl pyrrolidone) is a synthetic polymer with non-toxic, bio-inert and hydrophilic characteristics, making it a promising option for drug delivery applications in the pharmaceutical industry (Kumar, 2014). PVP is a binder utilized in many pharmaceutical tablets and PVP complexes are used in a variety of goods such as solutions, ointment, pessaries, liquid soaps and surgical scrubs (Karpuraranjith *et al*, 2017). In polar solvents such as alcohol, water, and others, PVP is highly soluble. It also has a high glass transition temperature ( $T_g$ ) and good environmental, thermal and mechanical stability. Another advantage of PVP is that it forms a thermally cross-connected polymer chain, which gives it exceptional thermal stability and great mechanical strength in mix composites. Poly(vinyl pyrrolidone) (PVP) was chosen as another polymer to construct polymer blends because of its outstanding properties (Sundaramahalingam *et al*, 2019).

## MATERIALS AND METHODS

### Polymer blend preparation

The polymer blend was prepared by solvent casting method. Chitosan solution was made by dissolving it in a 2 percent aqueous acetic acid solution and stirring it at room temperature. PVA and PVP were dissolved in hot water to form 5 wt% polymer solutions.

### Preparation of Ch/PVA/PVP-Ag, Zn nanocomposites

A 100 mg dried PVA/PVP/Ch mix was put in 50 mL of Ag and Zn solution at a concentration of 250 mg/L and sonicated for 1.5 hours to electrostatically bind Ag and Zn nano metals in the blend matrix.

## RESULTS AND DISCUSSION

### FT-IR analysis for polymers and polymer blend

From Fig. 1 for PVA, the bands at about 3369 and 1649 $\text{cm}^{-1}$  are assigned to stretching and bending vibration for hydroxyl group (Li *et al*, 2000). The band 2953 $\text{cm}^{-1}$  corresponding to asymmetric stretching vibration for ( $\text{CH}_2$ ) group. The band about 1151 $\text{cm}^{-1}$  for the acetyl groups (C–O) stretching vibration on the PVA backbone (Abdelaziz *et al*, 2007; Laot *et al*, 1999). The peak at 3392  $\text{cm}^{-1}$  in the FTIR spectrum of PVP (Fig. 2) shows O-H stretching. The peaks at 2923 and 1623  $\text{cm}^{-1}$ , respectively, showed the presence of asymmetric  $\text{CH}_2$  and C-O stretching. At 1457  $\text{cm}^{-1}$  and 1383  $\text{cm}^{-1}$ , respectively, C-H bending and  $\text{CH}_2$  wagging were detected (Rahma *et al*, 2016). The FTIR spectra for pure Chitosan (Fig. 3) can be assigned: Broad band at 3389 $\text{cm}^{-1}$

for stretching vibrations of (N–H and O–H) groups, 2953 $\text{cm}^{-1}$  ( $\text{CH}_3$  symmetric stretch), 1650 $\text{cm}^{-1}$  (C=O stretching vibration), 1458 $\text{cm}^{-1}$  (C–N stretching vibration), 1377 $\text{cm}^{-1}$  ( $\text{CH}_3$  bending vibration), 1153 $\text{cm}^{-1}$  (C–O–C bending vibration). For the prepared polymer blend. Fig. 4 showed a broad band around 3417 $\text{cm}^{-1}$  attributed to stretching vibration of hydroxyl group (–OH) of PVA and the secondary amide (–NH) of chitosan. The band at around 1046 $\text{cm}^{-1}$  indicates the presence of (OH) hydroxyl group with polymeric association and (–NH) a secondary amide. Also band appeared at 1452 $\text{cm}^{-1}$  assigned to pyridine ring (C=N).

### Scanning electron microscope studies (SEM)

The SEM micrograph for PVA/ PVP/Ch polymer blend and nanocomposite loaded with silver and zinc nanoparticles are represented in Figs. 5-7. The surface seems to be porous with some inclusions. Nanoparticles with an average size of 69nm for silver and 72nm for zinc particles are found in a homogeneous distribution over the matrix's surface.

### Thermal analysis

The thermo gravimetric (DSCD TGA) for PVA/ PVP/Chitosan polymer blend and its nano composite has been measured in temperature, which ranges between 25°C and 600°C with a constant rate which is equal to 10°C  $\text{min}^{-1}$ .

TGA curve of PVA/PVP/Chitosan polymer blend (B1), Fig. 8 illustrated four stages of a sequence mass loss, the first stage with mass loss (-8.935%) of volatile compounds. The second-step with weight loss approximately (-24.85%), the third stage with weight loss of approximately (-55.47%), the fourth stage with weight loss of approximately (-6.528%) for the chain decomposition. DSC curve in the Fig. 9 for polymer blend showed a  $T_g$  of (118.19°C). Peak at (446.81°C) regarding to the polymer melting  $T_m$ .

The TGA curve of PVA/PVP/Chitosan -Ag nanocomposite (NC-Ag), Fig. 9 illustrated four stages of a sequence mass loss, the first stage with mass loss (-10.18%) of volatile compounds. The second stage with weight loss approximately (-23.71%), the third stage with weight loss of approximately (-60.29%), the fourth stage with weight loss of approximately (-4.151%) for the chain decomposition. DSC curve in the Fig. 9 for polymer nanocomposite showed a  $T_g$  of (98.04°C), peak regarding to the Crystalline temperature point  $T_c$  at (351.98°C). Peak at (444.63°C) regarding to the polymer melting  $T_m$ .

The TGA curve of PVA/PVP/Chitosan -Zn nanocomposite (NC-Zn), Fig. 10 illustrated four stages of a sequence mass loss, the first stage with mass loss (-

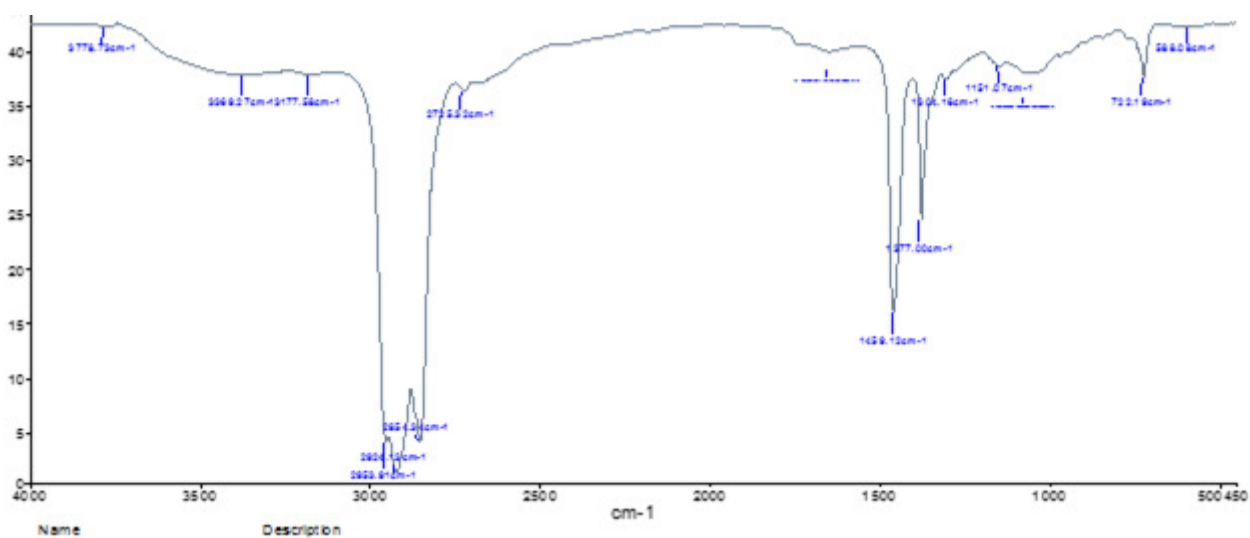


Fig. 1 : FT-IR spectrum PVA.

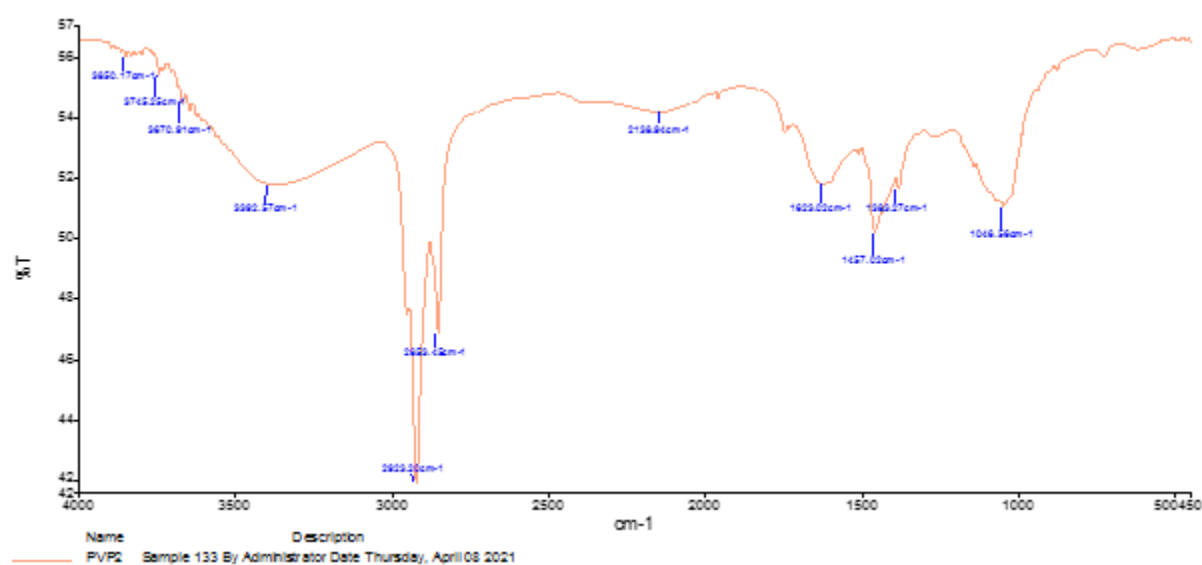


Fig. 2 : FT-IR spectrum PVP.

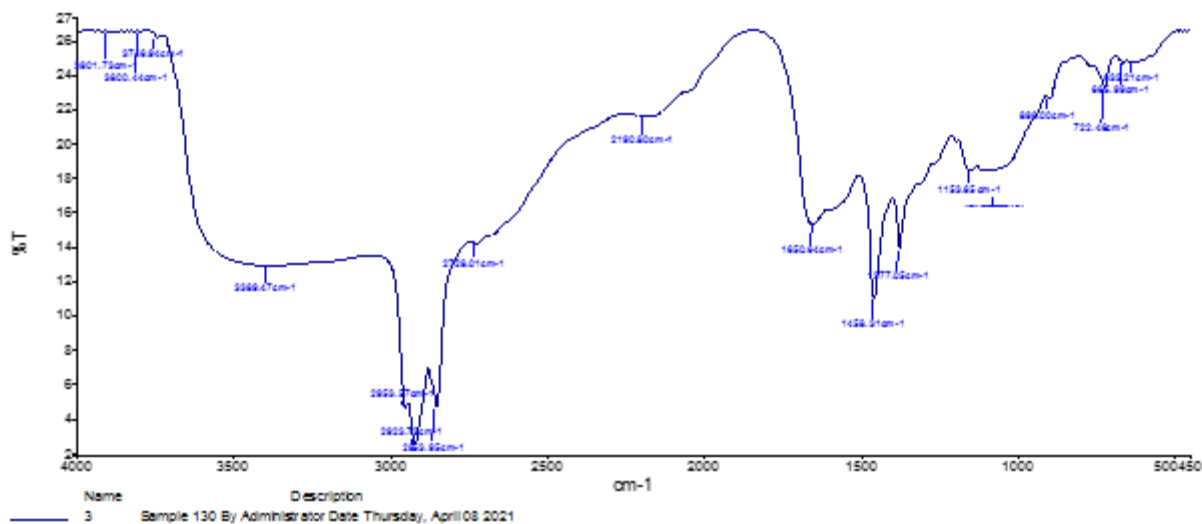


Fig. 3 : FT-IR spectrum Chitosan.

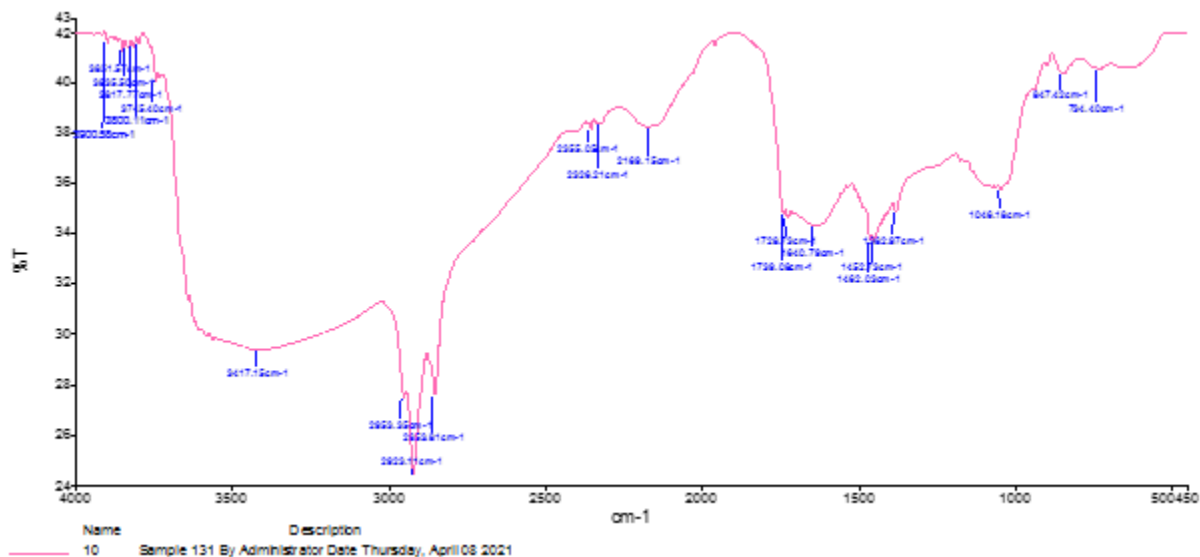


Fig. 4 : FT-IR spectrum PVA/PVP/Ch Blend.

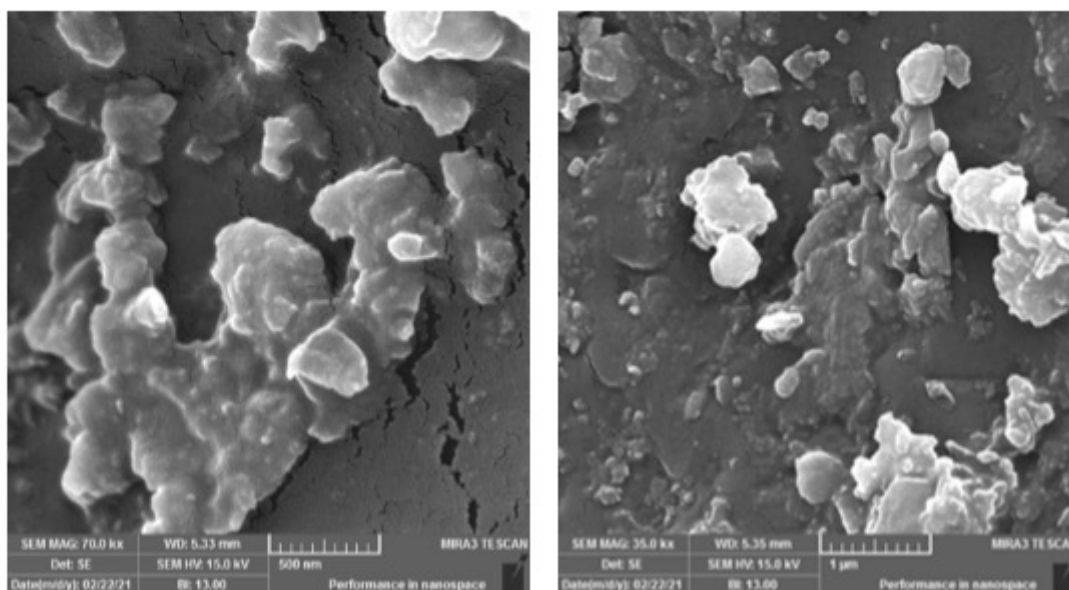


Fig. 5 : The SEM Images of blend B1.

10.61%) of volatile compounds. The second stage with weight loss approximately (-23.81%), the third stage with weight loss of approximately (-56.87%), the fourth stage with weight loss of approximately (-5.174%) for the chain decomposition. DSC curve in the Fig. 10 for polymer nanocomposite showed a Tg of (142.19°C), peak regarding to the Crystalline temperature point Tc at (311.84°C). Peak at (445.11°C) regarding to the polymer melting Tm.

### Biological activity

The biological activity of the polymer blend of PVA/PVP/Chitosan Ag and Zn nano composite were tested against six types of pathogenic bacteria using Diffusion inhibition method. The results of antimicrobial activity are represented in Table 1. When compared to Ag and Zn nanocomposites, the ternary mix (PVA/PVP/Chitosan)

that was employed as a control matrix demonstrated minimal antibacterial activity. It is clear from Table 1 that all tested compounds exhibit good antimicrobial activities.

For Ag nanocomposite the silver exhibits antibacterial property, which leads to biomedical applications. Silver's antibacterial action is based on Ag<sup>+</sup>, which binds tightly to electron donor groups in microbial cell wall such as sulfur, oxygen, or nitrogen. Silver ions work by displacing other important metal ions like Ca<sup>2+</sup> and Zn<sup>2+</sup> (Boomi *et al*, 2013). Silver nanoparticles have complex impacts on bacterial cells (Kim *et al*, 2011). However, the impact of silver nanoparticles on bacterial cells is mediated by a variety of ways (Prabhu *et al*, 2012). The following are some of the mechanisms that have been summarized and presented: I the capacity of silver nanoparticles to bind

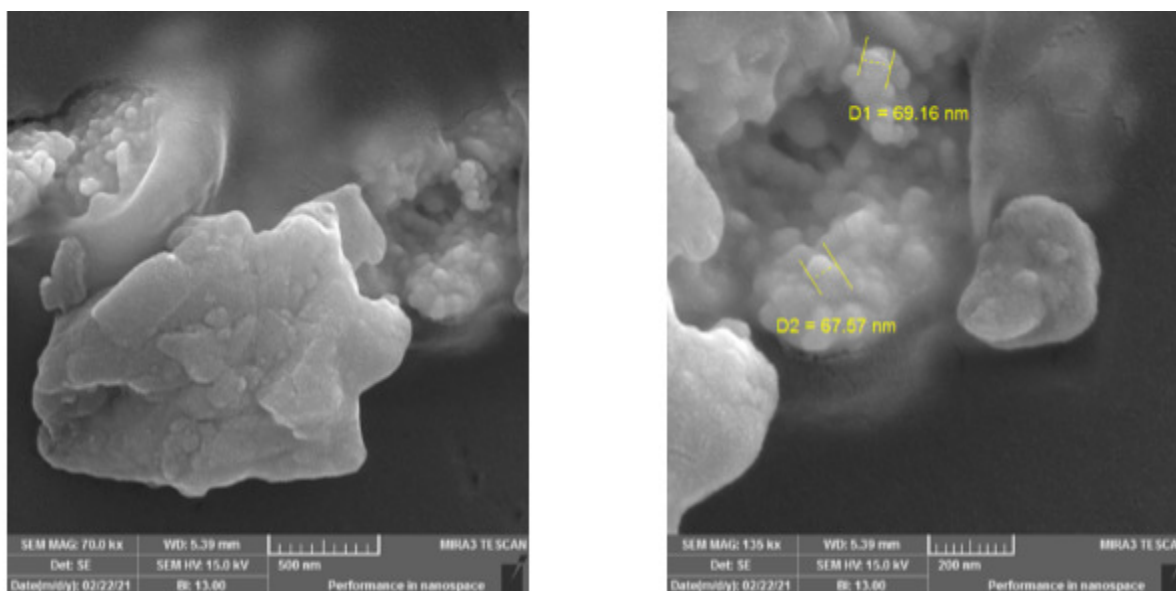


Fig. 6 : The SEM Images of Nanocomposite B1-Ag.

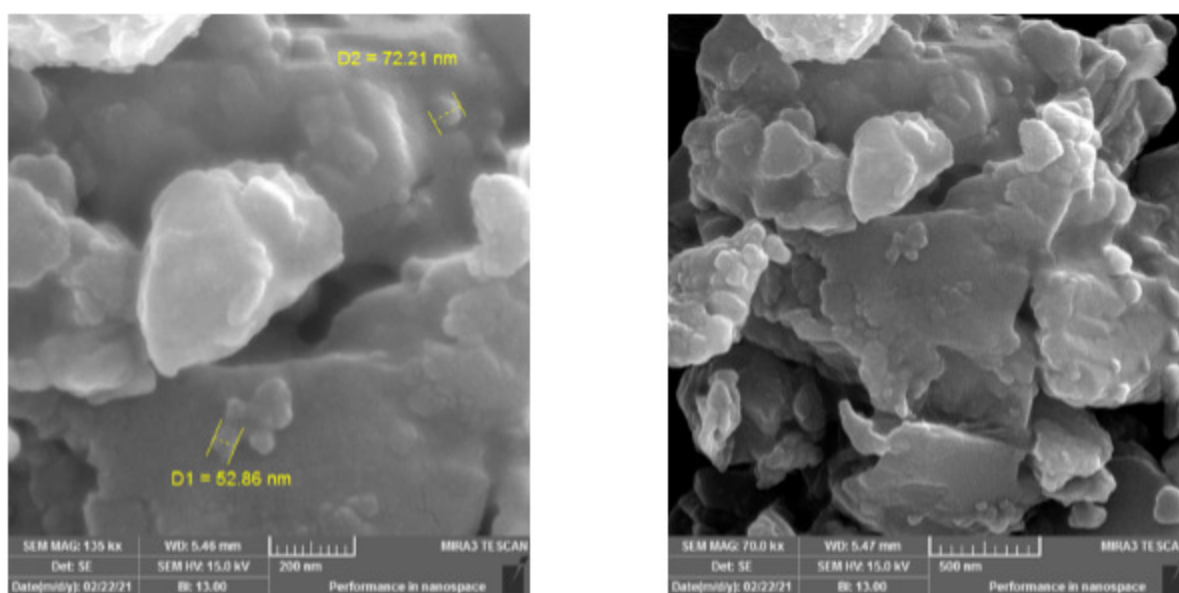


Fig. 7 : The SEM Image of Nanocomposite B1-Zn.

Table 1 : Antimicrobial activity of Polymer blend and Ag, Zn-nanocomposite on the bacterial and fungal isolates by well diffusion test.

Test	<i>C. albicans</i> Inhibition zone(mm)	<i>S.aureus</i> Inhibition zone (mm)	<i>E.faecalis</i> Inhibition zone (mm)	<i>E.coli</i> Inhibition zone (mm)	<i>S. typhi</i> Inhibition zone (mm)	<i>P. aeruginosa</i> Inhibition zone (mm)	<i>Bacillus cereus</i> Inhibition zone(mm)
B1	12	11	12	14	9	-	-
B2	13	9	11	12	11	-	-
B3	11	-	12	17	10	-	-
B1- Ag	22	21	18	26	21	18	22
B2- Ag	18	15	17	22	18	16	21
B3- Ag	15	13	15	19	16	11	20
B1- Zn	21	19	18	12	11	12	16
B2 - Zn	22	16	23	13	10	10	15
B3 - Zn	19	16	18	12	13	12	15



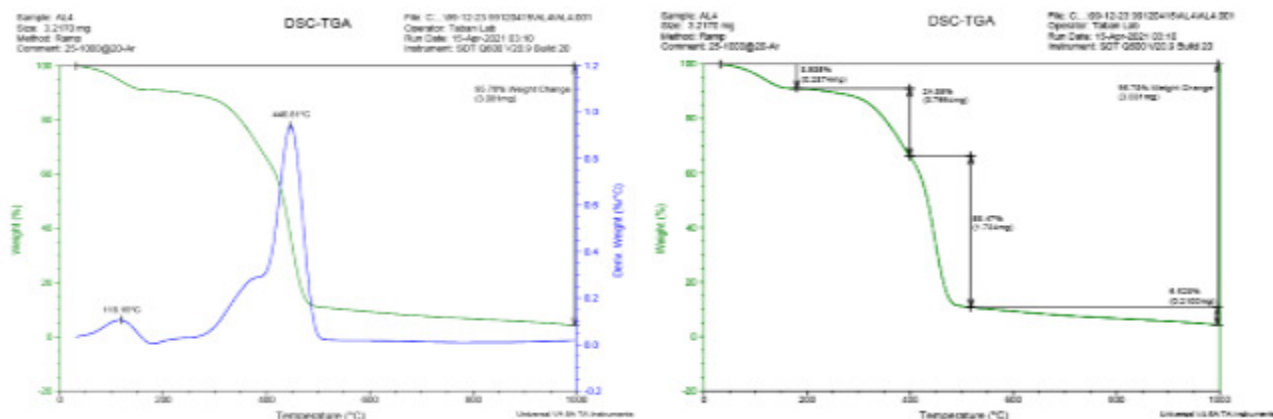


Fig. 8 : Thermal analysis of blend B1.

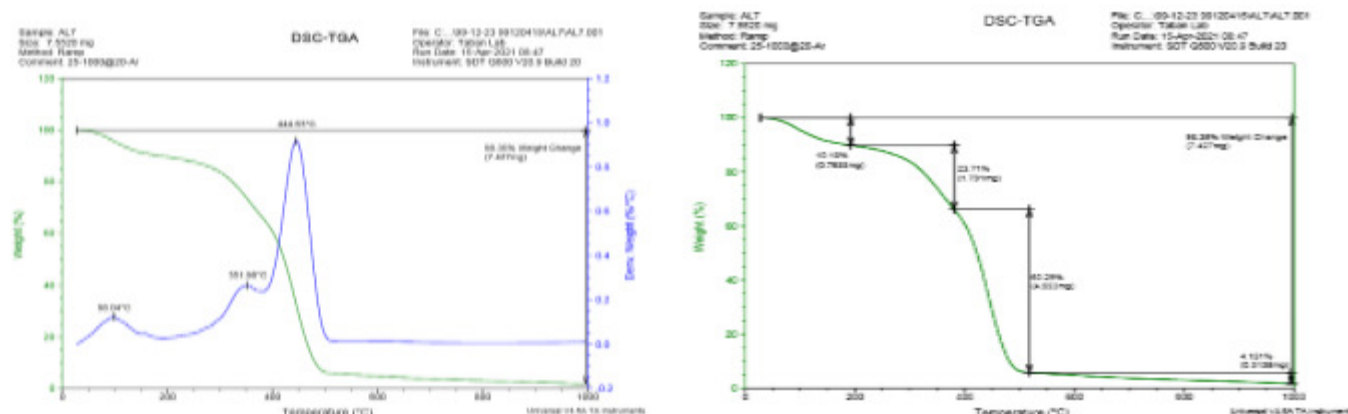


Fig. 9 : Thermal analysis of Nanocomposite B1Ag.

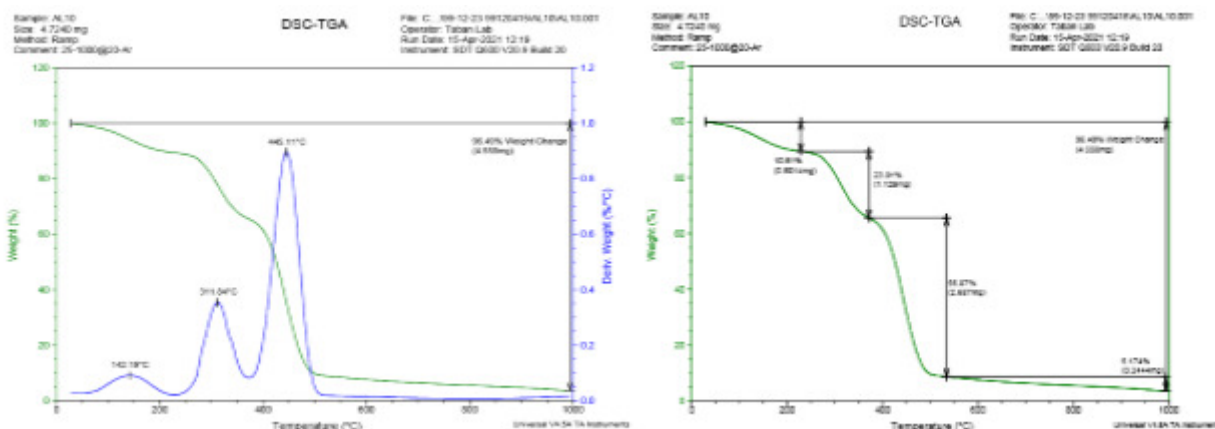


Fig. 10 : Thermal analysis of Nanocomposite B1Zn.

to and enter the bacterial cell wall (Sondi *et al.*, 2004), (ii) the production of free radicals by silver nanoparticles, which may damage and porous the cell membrane (Danilcauk *et al.*, 2006), (iii) nanoparticles can release silver ions, which can bind and inactivate the thiol groups of several important enzymes (Brian Y Feng *et al.*, 2008), and (iv) nanoparticles can alter signal transduction in bacteria, preventing bacteria from growing (Shrivastava *et al.*, 2007).

For Zn nanocomposite several mechanisms have described Zn antimicrobial activity, including cell damage

through their interaction with the microorganisms and the reactive oxygen species (ROS) formation by the activation of this metal (Ghaffari-Moghaddam *et al.*, 2014; Dicastillo *et al.*, 2020).

## REFERENCES

- Abdelaziz M and Abdelrazek E M (2007) Effect of dopant mixture on structural, optical and electron spin resonance properties of polyvinyl alcohol. *Physica B: Condensed Matter* **390**(1-2), 1-9.
- Aranaz I, Mengibar M, Harris R, Paños I, Miralles B, Acosta N and Heras Á (2009) Functional characterization of chitin and chitosan. *Curr. Chem. Biol.* **3**(2), 203-230.

- Arefian M, Hojjati M, Tajzad I, Mokhtarzade A, Mazhar M and Jamavari A (2020) A review of Polyvinyl alcohol/Carboxy methyl cellulose (PVA/CMC) composites for various applications. *J. Composites and Compounds* **2**(3), 69-76.
- Aytumur A, Koçyiđit S and Uslu Ý (2013) Synthesis and characterization of poly (vinyl alcohol)/poly (vinyl pyrrolidone)-iodine nanofibers with poloxamer 188 and chitosan. *Polymer-Plastics Technology and Engineering* **52**(7), 661-666.
- Boomi P, Prabu H G and Mathiyarasu J (2013) Synthesis and characterization of polyaniline/Ag-Ptnanocomposite for improved antibacterial activity. *Colloids and Surfaces B: Biointerfaces* **103**, 9-14.
- Brian Y Feng, Brandon H Toyama and Holger Wille (2008) Small-molecule aggregates inhibit amyloid polymerization. *Nat. Chem. Biol.* **4**, 197-199.
- Cheung R C F, Ng T B, Wong J H and Chan W Y (2015) Chitosan: an update on potential biomedical and pharmaceutical applications. *Marine Drugs* **13**(8), 5156-5186.
- Danilcauk *et al* (2006) Conduction electron spin resonance of small silver particles. *Spectrochimica Acta Part A Molecular and Biomolecular Spectroscopy* **63**(1), 189-191.
- Dicastillo C L D, Vidal C P, Falcó I, Sánchez G, Márquez P and Escrig J (2020) Antimicrobial bilayer nanocomposites based on the incorporation of as-synthesized hollow zinc oxide nanotubes. *Nanomaterials* **10**(3), 503.
- Garavand F, Cacciotti I, Vahedikia N, Rehman A, Tarhan Ö, Akbari-Alavijeh S and Jafari S M (2020) A comprehensive review on the nanocomposites loaded with chitosan nanoparticles for food packaging. *Critical Rev. Food Sci. Nutrition* **2020**, 1-34.
- Ghaffari-Moghaddam M and Eslahi H (2014) Synthesis, characterization and antibacterial properties of a novel nanocomposite based on polyaniline/polyvinyl alcohol/Ag. *Arabian J. Chem.* **7**(5), 846-855.
- Joseph B, Krishnan S, Sagarika V K, Tharayil A, Kalarikkal N and Thomas S (2020) Bionanocomposites as industrial materials, current and future perspectives: a review. *Emergent Materials* 1-15.
- Jou C H (2011) Antibacterial activity and cytocompatibility of chitosan-N-hydroxy-2, 3-propyl-N methyl-N, N-diallylammonium methyl sulfate. *Colloids and Surfaces B: Biointerfaces* **88**(1), 448-454.
- Kassem A, Ayoub G M and Malaeb L (2019) Antibacterial activity of chitosan nano-composites and carbon nanotubes: A review. *Sci. Total Environ.* **668**, 566-576.
- Kim S H, Lee H S, Ryu D S, Choi S J and Lee D S (2011) Antibacterial activity of silver-nanoparticles against *Staphylococcus aureus* and *Escherichia coli*. *Microbiol. Biotech. Lett.* **39**(1), 77-85.
- Kumar D, Gihar S, Shrivash M K, Kumar P and Kundu P P (2020) A review on the synthesis of graft copolymers of chitosan and their potential applications. *Int. J. Biol. Macromolecules* **163**, 2097-2112.
- Kumar G P, Phani A R, Prasad R G S V, Sanganal J S, Manali N, Gupta R and Raju D B (2014) Polyvinylpyrrolidone oral films of enrofloxacin: Film characterization and drug release. *Int. J. Pharmaceutics* **471**(1-2), 146-152.
- Laot C M, Marand E and Oyama H T (1999) Spectroscopic characterization of molecular interdiffusion at a poly (vinyl pyrrolidone)/vinyl ester interface. *Polymer* **40**(5), 1095-1108.
- Li X, Goh S H, Lai Y H and Wee A T S (2000) Miscibility of carboxyl-containing polysiloxane/poly (vinylpyridine) blends. *Polymer* **41**(17), 6563-6571.
- Martins A F, Facchi S P, Follmann H D, Pereira A G, Rubira A F and Muniz E C (2014) Antimicrobial activity of chitosan derivatives containing N-quaternized moieties in its backbone: a review. *Int. J. Mol. Sci.* **15**(11), 20800-20832.
- Parida U K, Nayak A K, Binhani B K and Nayak P L (2011) Synthesis and characterization of chitosan-polyvinyl alcohol blended with cloisite 30B for controlled release of the anticancer drug curcumin. *J. Biomaterials and Nanobiotech.* **2**(04), 414.
- Prabhu S and Poulouse E K (2012) Silver nanoparticles: mechanism of antimicrobial action, synthesis, medical applications, and toxicity effects. *Int. nano Lett.* **2**(1), 1-10.
- Qin Y, Li P and Guo Z (2020) Cationic chitosan derivatives as potential antifungals: A review of structural optimization and applications. *Carbohydrate Polymers* **236**, 116002.
- Raafat D and Sahl H G (2009) Chitosan and its antimicrobial potential—a critical literature survey. *Microbial Biotech.* **2**(2), 186-201.
- Rahma A, Munir M M, Khairurrijal Prasetyo A, Suendo V and Rachmawati H (2016) Intermolecular interactions and the release pattern of *Electrospun curcumin* Polyvinyl (Pyrrolidone) Fiber. *Biol. Pharmaceut. Bull.* **39**(2), 163-173.
- Rezkita F, Sarasati A, Wijaya F N, Nugraha A P, Hendrijantini N, Ridwan R D, Ramadhani N F, Fadholly A, Ansori A N M and Kuncoroningrat Susilo R J (2020) Chitosan scaffold, concentrated growth factor and gingival mesenchymal stem cells as the osteoporotic jawbone therapy: A review. *Biochem. Cell. Arch.* **20**, 2913-2919.
- ScrubsKarpuraranjith M and Thambidurai S (2017) Chitosan/zinc oxide-polyvinylpyrrolidone (CS/ZnO-PVP) nanocomposite for better thermal and antibacterial activity. *Int. J. Biol. Macromolecules* **104**, 1753-1761.
- Shrivastava S, Bera T, Roy A, Singh G, Ramachandrarao P and Dash D (2007) Characterization of enhanced antibacterial effects of novel silver nanoparticles. *Nanotechnology* **18**(22), 225103.
- Sondi I and Salopek-Sondi B (2004) Silver nanoparticles as antimicrobial agent: a case study on E. coli as a model for Gram-negative bacteria. *J. Colloid and Interface Sci.* **275**(1), 177-182.
- Sundaramahalingam K, Muthuvinayagam M and Nallamuthu N (2019) AC impedance analysis of lithium ion based PEO: PVP solid polymer blend electrolytes. *Polymer Science, Series A* **61**(5), 565-576.
- Wei Y, Li C, Zhang L, Dai L, Yang S, Liu J and Gao Y (2020) Influence of calcium ions on the stability, microstructure and in vitro digestion fate of zein-propylene glycol alginate-tea saponin ternary complex particles for the delivery of resveratrol. *Food Hydrocolloids* **106**, 105886.